THE EFFECTS OF CADMIUM AND COENZYME Q_{10} ON ANTIOXIDANT DEFENCE ENZYME ACTIVITIES IN THE BLOOD OF RATS

SLADJAN Z. PAVLOVIĆ^{1*}, BRANKA I. OGNJANOVIĆ², ANDRAŠ Š. ŠTAJN², RADOSLAV V. ŽIKIĆ², ZORICA S. SAIČIĆ¹, VOJISLAV M. PETROVIĆ¹

Institute for Biological Research "Siniša Stanković", Department of Physiology, Belgrade, Serbia, Yugoslavia
²Faculty of Sciences, Institute of Biology, University of Kragujevac, Kragujevac, Serbia, Yugoslavia

The purpose of our study was to investigate the effects of cadmium (Cd) and coenzyme Q_{10} (Co Q_{10}) on the activities of copper, zinc-containing superoxide dismutase (CuZn SOD), catalase (CAT) and glutathione peroxidase (GSH-Px) in red blood cells (RBC) as well as glutathione-S-transferase (GST) in the plasma of male two months old *Wistar albino* rats. The animals were divided in five experimental groups as follows: (1) control (C), (2) Cd (17 mg/day/kg body mass + 100 μ l olive oil, every fifth day), (3) Co Q_{10} (16/mg/kg body mass, dissolved in olive oil, every fifth day), (4) Cd+Co Q_{10} (in the above mentioned amounts) and (5) olive oil (o. oil, 100 μ l i.m., every fifth day) and treated in the course of 30 days. After the treatment, the animals were sacrificed and blood samples were prepared for the analysis. Our results show that Cd administered with olive oil did not exhibit toxic effects on CuZn SOD, CAT and GSH-Px activities in RBC as well as GST activity in the plasma of rats. At the same time, Co Q_{10} by quenching the free oxygen radicals and by inhibiting lipid peroxidation may improve the antioxidant defence enzyme activities in the blood of rats. Our investigations also show that olive oil exhibit some protective effects on CuZn SOD, CAT and GSH-Px activities in RBC and GST activity in the plasma of rats treated with Cd.

INTRODUCTION

Cadmium (Cd) has been recognized as one of the most toxic environmental and industrial pollutants (Yiin, Chern, Sheu, Tseng & Lin, 1999). Major routes of Cd intake into the organism are respiratory and gastrointestinal tract (Gupta, Murthy, Thauur, Dudley & Chandra, 1990). After absorption, Cd penetrate in the bloodstream and binds to the erythrocyte membranes and plasma albumin (Waalkes & Oberdorster, 1990). In the blood and tissues Cd induces formation of metallothioneins which represent the complexes of Cd and lowmolecular mass proteins (Simpkins, Lloyd & Balderman, 1998). Cd stimulates formation of reactive oxygen species and influences oxidative damage in erythrocytes resulting in loss of membrane function (Sarkar, Yadav & Bhatnagar, 1997). Cadmium induces also onset of anemia, decreases hemoglobin concentration, lowers the hematocrit value as well as decreases blood iron level (Kostić, Ognjanović, Dimitrijević, Žikić, Štajn, Rosić &

Živković, 1993). A variety of accompanying changes in antioxidant defence enzymes were reported (Shukla & Chandra, 1989).

Coenzyme Q (CoQ) is the only lipid-soluble antioxidant which is normally synthesized by the organism (Lenaz, Bovina, Formiggini & Parenti-Castelli, 1999). CoO is present in almost all living organisms where it exhibits antioxidant properties and also represents an obligatory component of the mitochondrial respiratory chain (Ernster & Dallner, 1995). Many investigations demonstrate the antioxidant capacity of reduced form of CoQ10 (CoQ10H2) over the oxidized form (Mellors & Tappel, 1966). CoQ10 inhibits the process of lipid peroxidation (Ernster & Dallner, 1995), regenerates the active form of vitamin E from the vitamin E radical (Kozlov, Gille, Staniek & Nohl, 1999) and stabilizes the extracellular ascorbate in the organism (Gomez-Diaz, Rodtiguez-Aguilera, Barroso, Villalba, Navarro, Crane & Navas, 1997). CoQ10 can protect organism from oxidative stress induced by various toxic agents (Beyer, 1988).

^{*}Correspondence to: Sladjan Z. Pavlović, M. Sc., Research Assistant, Institute for Biological Research "Siniša Stanković" Department of Physiology, 29. Novembra 142, 11060 Belgrade, Serbia, Yugoslavia; phone: (+381) 11 764 422, ext. 141; fax: (+381) 11 761 433, e-mail: sladjan@ibiss.bg.ac.yu

Oxidative damage induced by carbon tetrachloride was decreased by CoQ₁₀ administration (Bertelli, Giovannini, Bertelli, Maltinti, Scalori & Romano, 1986). Coenzyme Q₁₀ inhibits also lipid peroxidation induced by carbon tetrachloride and ethanol (Beyer, 1988). CoQ₁₀ has a large number of clinical applications, especially in the treatment of cardiovascular and neurologic diseases (Mortensen, Bouchelouche, Maratsu & Folkers, 1986; Zierz, Jahns & Jerusalem, 1989).

The aim of this study was to evaluate the blood antioxidant defence enzyme activities in rats chronically treated with Cd, CoQ₁₀, concomitantly treated with Cd+CoQ₁₀ and olive oil. After 30 days of exposure the activities of copper, zinc-containing superoxide dismutase (CuZn SOD, EC 1.15.1.1), catalase (CAT, EC 1.11.1.6) and glutathione peroxidase (GSH-Px, EC 1.11.1.9) in red blood cells (RBC) as well as glutathione-Stransferase (GST, EC 2.5.1.18) in the plasma of rats were estimated.

MATERIALS AND METHODS

The experiments were carried out with male, 60 days old Wistar albino rats, weighing 190 ± 20 g at the beginning of experiments. The animals were haused in individual cages at 21 ± 1°C and exposed to 12hr light-12hr dark cycle. The rats were fed chow pellets (Veterinarski Zavod, Zemun, Yugoslavia) and drank tap water ad libitum. The animals were divided in five experimental groups and treated in the course of 30 days. First group of animals was control (C, drinking tap water). Second group was treated with cadmium (Cd, 200 mg CdCl₂ × 5H₂O in drinking water during 30 days + 100 µl of olive oil, i.m., every fifth day). Third group was treated with Coenzyme Q10 (CoQ10, 20 mg CoQ10 dissolved in olive oil, i.m., every fifth day, drinking tap water). Fourth group was treated concomitantly with cadmium and Coenzyme Q10 (Cd+CoQ10, 200 mg CdCl2 × 5H2O in drinking water during 30 days + 20 mg CoQ10, i.m. every fifth day). Fifth group was treated with olive oil (o. oil, 100 µl), i.m., every fifth day (drinking tap water). The average intake of 17 mg Cd/(day × kg body mass) was calculated from the water consumed during the 30 days of treatment. The average intake of CoQ10 was 16 mg/kg body mass every fifth day. Every group consisted of 7 animals. After the treatment the animals were sacrificed by decapitation between 8 and 10 am in order to avoid any possible circadian changes in antioxidant defence enzyme activities.

Aliquots of fresh blood were taken immediately after exsanguination, collected in heparinized test tubes (1000 IU of heparin) and centrifuged (5000 rpm) for the separation of plasma and blood cells. RBC was washed three times with 3 vol. of cold 155 mmol/l NaCl. Haemolysates containing about 50 g Hb/l were prepared according to McCord and Fridovich (1969) and used for the determination of CAT and GSH-Px activities. Measurement of CuZn SOD activity was conducted in the haemolysates of washed RBC, in which Hb was previously removed by the method of Tsuchihashi (1923). CuZn SOD activity was measured by the epinephrine method (Misra & Fridovich, 1972) based on the capacity of SOD to inhibit autooxidation of adrenaline to adrenochrome. One unit of SOD activity was defined as the amount of the enzyme causing 50% inhibition of the autooxidation of adrenaline. CAT activity was assayed as suggested by Beutler (1982) and expressed as µmol H₂O₂/(min × g Hb). The activity of GSH-Px was evaluated by the method of Maral, Puget and Michelson (1977) based on the measurement of nicotine amide dinucleotide phosphate (NADPH) consumption, and expressed as nmol NADPH/ (min × g Hb). For the determination of GST activity in the plasma 1-chloro-2,4-dinitrobenzene (CDNB) was used as a substrate (Habig, Pubst & Jakoby, 1974). The method is based on the reaction of CDNB with -SH groups of glutathione catalyzed by GST contained in the samples, and one unit of GST activity was defined as nmol GSH/(min × ml) of plasma.

Data are given as means \pm SE. All obtained results were compared with respect to control animals (C) as well as to the animals treated with olive oil (o. oil) in order to diffentiate the effects of CoQ_{10} from the effects of o. oil. Statistical analysis of the results was based on the Student's paired "t" test considering the significance at a level of P < 0.05 (Hoel, 1966).

RESULTS

In comparison to the control animals the activity of CuZn SOD in RBC was significantly decreased (A: P < 0.05) only in rats treated with olive oil (Fig. 1). At the same time in relation to the animals treated with olive oil, CuZn SOD activity was significantly increased in rats treated with Cd and CoQ_{10} (a: P < 0.05) as well as in animals concomitantly treated with Cd + CoQ_{10} (c: P < 0.01).

The activity of CAT in RBC is presented in Fig. 2. The obtained results show that, with respect to

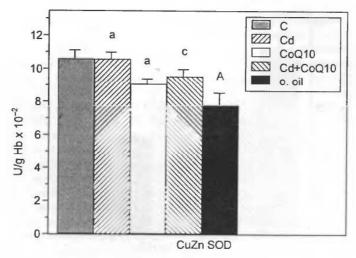


Fig. 1. The activity of copper zinc containing superoxide dismutase (CuZn SOD) expressed in U/g Hb in red blood cells (RBC) of control animals (C), treated with cadmium (Cd, 17 mg/(day × kg body mass) in drinking water + 100 μl olive oil, every fifth day, i.m.), treated with coenzyme Q₁₀ (CoQ₁₀, 16 mg/(kg × every fifth day), i.m., dissolved in olive oil), concomitantly treated with cadmium and coenzyme Q₁₀ (Cd + CoQ₁₀, 17 mg/(day × kg body mass) in drinking water + 16 mg/(kg × every fifth day), i.m., dissolved in olive oil) and animals treated with olive oil (o. oil, 100 μl every fifth day, i.m.). The values are means ± SE from seven animals in each group. The results were compared in respect to the control animals (C) as well as to the animals treated with olive oil (o. oil). Significance from C: ^AP < 0.05. Significance from o. oil: ^aP < 0.05; ^cP < 0.01

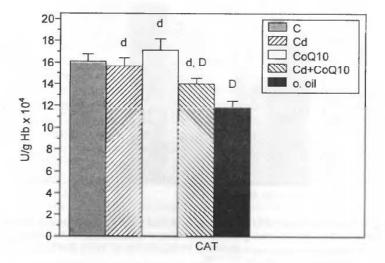


Fig. 2. Catalase (CAT) activity expressed in U/g Hb in RBC of animals. The same experimental groups as in Fig. 1. The values are means \pm SE from seven animals in each group. The results were compared in respect to the control animals (C), as well as to the animals treated with olive oil (o. oil). Significance from C: $^{D}P < 0.005$. Significance from 0. oil: $^{d}P < 0.005$

the control rats, CAT activity was significantly decreased in animals concomitantly treated with Cd + CoQ_{10} and in animals treated with olive oil (D: P < 0.005). In relation to the animals treated with olive oil, CAT activity was significantly increased in all experimental groups of animals:

those treated with Cd, CoQ_{10} and those treated concomitantly with Cd + CoQ_{10} (d: P < 0.005).

The GSH-Px activity in RBC of rats is depicted in Fig. 3 with respect to the control animals GSH-Px activity was significantly decreased in animals treated with CoQ_{10} and olive oil (A: P < 0.05). On

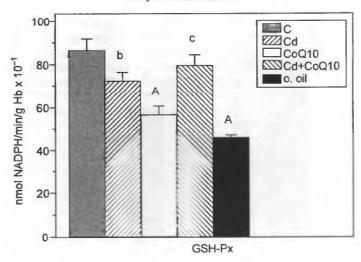


Fig. 3. The activity of glutathione peroxidase (GSH-Px) expressed in nmol NADPH/(min \times g Hb) in RBC of animals. The same experimental groups as in preceding Figs. The values are means \pm SE from seven animals in each group. The results were compared in respect to the control animals (C), as well as to the animals treated with olive oil (o. oil). Significance from C: $^{4}P < 0.05$. Significance from o. oil: $^{6}P < 0.02$; $^{6}P < 0.01$

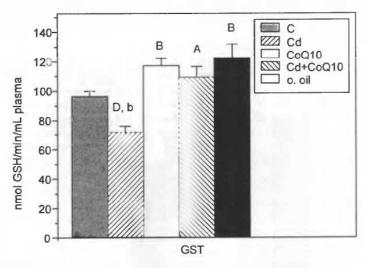


Fig. 4. Plasma glutathione-S-transferase (GST) activity expressed in nmol GSH/(min \times ml plasma). The same experimental groups as in preceding Figs. The values are means \pm SE from seven animals in each group. The results were compared in respect to the control animals (C), as well as to the animals treated with olive oil (o. oil). Significance from C: $^{A}P < 0.05$; $^{B}P < 0.02$; $^{D}P < 0.005$. Significance from o. oil: $^{b}P < 0.02$

the other hand RBC GSH-Px activity was significantly increased in animals treated with Cd (b: P < 0.02) and concomitantly treated with Cd + CoQ₁₀ (c: P < 0.01) in comparison to the rats treated with olive oil.

The activity of GST in the plasma was presented in Fig. 4 with respect to the controls the plasma GST activity was significantly decreased in rats treated with Cd (D: P < 0.005) and significantly increased in animals treated with CoQ_{10} (B: P < 0.02), concomitantly treated with Cd + CoQ_{10} (A: P < 0.05) and treated with olive oil (B: P < 0.02). The activity of plasma GST was significantly decreased only in animals treated with Cd (b: P < 0.02) in relation to the rats treated with olive oil.

DISCUSSION

During the evolution mammalian cells have developed protective mechanisms to minimize injurious events that result from normal oxidative products of cellular metabolism and toxic chemicals. The antioxidant defence system is species- and organ-specific (Petrović, Saičić, Spasić, Radojičić & Buzadžić, 1991). The extent of the oxidant damage to the erythrocytes could be used as a suitable biomarker for the estimation of general oxidant damage in the organisms (Saičić, Simović, Korać, Blagojević, Buzadžić, Spasić & Petrović, 1993).

Many authors have reported that Cd administered alone leads to an increased production of superoxide anion radicals in RBC of rats (Singhal, Anderson & Meister, 1987; Kostić et al., 1993). It is reasonable to expect an increased activity of CuZn SOD in RBC of Cd treated animals as a biological response. However, in our experiments perorally administered Cd was combined with i.m. injections of olive oil every fifth day. As a consequence, the activity of CuZn SOD in RBC of animals was not changed in comparison to the control value. We propose that olive oil administered together with Cd diminished the toxic effects provoked by Cd when it was administered alone. Namely, olive oil contains many antioxidants, such as vitamin E, coenzyme Q and polyphenols, which have many beneficial effects against oxidative injuries caused by various toxicants (Quiles, Ramirez-Tortosa, Huertas, Ibanez, Gomez, Battino & Mataix, 1999). At the same time, in animals treated with olive oil the activity of CuZn SOD was even significantly lowered in comparison to the controls. Consequently, when compared to the animals treated with olive oil, the activity of this enzyme was significantly increased in animals treated with Cd and Cd + CoQ10.

Superoxide dimutase has a function to eliminate the toxic superoxide anion radicals forming hydrogen peroxide (H2O2) which represents a substrate for CAT and GSH-Px (McCord & Fridovich, 1969). It is well known that the activity of CAT is directly proportional to the substrate level assumed to be produced by SOD (Aebi, 1974). This could be an explanation for the unchanged activity of CAT in RBC of Cd-treated animals in relation to the control value. On the other hand, CAT activity was significantly decreased in RBC of animals concomitantly treated with Cd + CoQ10, as well as in animals treated with olive oil. Decreased activity of CAT in RBC of Cd + CoQ10 treated animals was accompanied by an unchanged activity of GSH-Px in relation to the controls. GSH-Px destroys H₂O₂ in erythrocytes more efficiently than CAT. Decreased activity of CAT in RBC of rats treated with olive oil is accompanied by decreased activity of SOD and smaller production of H₂O₂.

The main antioxidant function of CoQ10 is to prevent the formation of lipid peroxyl radicals (LOO) and to prevent both initiation and propagation of lipid peroxidation (Ernster & Dallner, 1995). At the same time, GSH-Px has a crucial role in detoxification of H2O2 and organic hydroperoxides (Halliwell & Gutteridge, 1999). By quenching organic oxygen-derived free radicals, CoQ10 protects GSH-Px and activity of this enzyme molecule in RBC of rats treated with CoQ10 was significantly decreased with respect to the control value. A decreased activity of GSH-Px was also obtained in RBC of rats treated with olive oil. Earlier investigations have shown that olive oil contains polyphenolic antioxidants such as (3,4-dihydroxyphenyl)-ethanol and (p-hydroxyphenyl)ethanol which by quenching free oxygen radicals may lower risk of reactive oxygen species-mediated injuries (Manna, Galletti, Cucciolla, Moltedo, Leone & Zappia, 1997). Our earlier investigations have shown that other antioxidants such as selenium significantly increased activity of GSH-Px in tissues of rats (Žikić, Štajn, Ognjanović, Saičić, Kostić, Pavlović & Petrović, 1998) and thus protected tissues from Cd toxicity. Obviously, this mechanism of protection is different from the mechanism by which CoQ10 protects tissues from oxidative damage, because Se is the constituent of the active site of GSH-Px (Stadtman, 1980).

The results of our investigations show that Cd induced a significant decrease of GST activity in the plasma of rats. Similar results were obtained in livers of rats treated by carbon tetrachloride (Takahashi, Sugimoto, Takahata, Okamoto & Kishi, 1996). After the treatment of rats with CoQ₁₀ the activity of GST was significantly elevated in respect to the controls as well as to the rats treated with Cd. Same authors (Takahashi *et al.*, 1996) were also found that CoQ₁₀ elevates the activity of GST and thus improve defence against oxidant injuries caused by carbon tetrachloride. Various antioxidants contained in olive oil also influence the elevation of GST activity in the plasma of rats.

CONCLUSIONS

From the presented results it may be concluded that Cd administered with olive oil did not exhibit toxic effects on CuZn SOD, CAT and GSH-Px activities in RBC as well as GST activity in the

plasma of rats. At the same time, CoQ₁₀ by quenching the free oxygen radicals and by inhibiting the lipid peroxidation may improve the antioxidant defence enzyme activities in the blood of rats. Our investigations also show that olive oil exhibits some protective effects on CuZn SOD, CAT and GSH-Px activities in RBC and GST activity in the plasma of rats treated with Cd.

Acknowledgments

The work reported here was supported by the Ministry of Science and Technology of Serbia, grant Nos 03E18 and 03E23.

REFERENCES

- Aebi H. (1974). Catalase, [In:] Methods in Enzymatic Analysis. Bergmeyer H. U. (ed.), p. 73, Academic Press, New York and London.
- Bertelli A., Giovannini I., Bertelli A. A., Maltinti G., Scalori V. & Romano M. R. (1986). Tissue concentrations of coenzyme Q in rats intoxicated by carbon tetrachloride. *Int. J. Tissue. React.*, 8, 343–346.
- Beutler E. (1982). Catalase, [In:] Red Cell Metabolism, a Manual of Biochemical Methods. Beutler E. (ed.), pp. 105-106, Grune and Stratton, New York, USA.
- Beyer R. E. (1988). Inhibition by coenzyme Q of ethanol- and carbon tetrachloride-stimulated lipid peroxidation in vivo and catalyzed by microsomal and mitochondrial systems. Free Radic. Biol. Med., 5, 397 –303.
- Ernster L. & Dallner G. (1995). Biochemical, physiological and medical aspects of ubiquinone function. Biochim. Biophys. Acta, 1271, 195–204.
- Gomez-Diaz C., Rodriguez-Aguilera J. C., Barroso M. P., Villalba J. M., Navarro F., Crane F. L. & Navas P. (1997). Antioxidant ascorbate is stabilized by NADH-coenzyme Q₁₀ reductase in the plasma membrane. J. Bioenerg. Biomembr., 29, 251–257.
- Gupta A., Murthy C. R., Thauur R. S., Dudley P. M. & Chandra V. S. (1990). Comparative neurotoxicity of cadmium in growing and adult rats after repeated administration. *Biochem. Int.*, 21, 97–105.
- Habig W. H., Pubst M. J. & Jakoby W. B. (1974). Glutathione S-transferase. J. Biol. Chem., 249, 7130 -7139.
- Halliwell B. & Gutteridge J. M. C. (1999). Free Radicals in Biology and Medicine, Third edition, Oxford University Press Inc., New York.
- Hoel P. G. (1966). Introduction to Mathematical Statistics. John Wiley and Sons, New York, pp. 402–403.
- Kostić M. M., Ognjanović B., Dimitrijević S., Žikić R. V., Štajn A., Rosić G. L. & Živković R. V. (1993). Cadmium-induced changes of antioxidant and metabolic status in red blood cells of rats: in vivo effects. Eur. J. Haematol., 51, 86–92.

- Kozlov A. V., Gille L., Staniek K. & Nohl H. (1999). Dihydrolipoic acid maintains ubiquinone in the antioxidant active form by two-electron reduction of ubiquinone and one-electron reduction of ubisemiquinone. Arch. Biochem. Biophys., 363, 148–154.
- Lenaz G., Bovina C., Formiggini G. & Parenti-Castelli G. (1999). Mitochondria, oxidative stress, and antioxidant defences. Acta Biochim. Pol., 46, 1–21.
- Manna C., Galletti P., Cucciolla V., Moltedo O., Leone A. & Zappia V. (1997). The protective effect of the olive oil polyphenol (3,4-dihydrohyphenyl)-ethanol counteracts reactive oxygen metabolite-induced toxicity in Caco-2 cells. J. Nutr., 127, 286–292.
- Maral J., Puget K. & Michelson A. M. (1977). Comparative study of superoxide dismutase, catalase and glutathione peroxidase levels in erythrocytes of different animals. *Biochem. Biophys. Res. Commun.*, 77, 1525–1535.
- McCord J. M. C. & Fridovich I. (1969). Superoxide dismutase. An enzymatic function for erythrocuprein (haemocuprein). J. Biol. Chem., 244, 6049–6056.
- Mellors A. & Tappel A. L. (1966). Quinones and quinols as inhibitors of lipid peroxidation. *Lipids*, 1, 282–284.
- Misra H. P. & Fridovich I. (1972). The role of the superoxide anion in the autooxidation of epinephrine and simple assay for superoxide dismutase. J. Biol. Chem., 247, 3170–3175.
- Mortensen S. A., Bouchelouche P., Maratsu K. & Folkers K. (1986). [In:] Clinical Decline and Relapse of Cardiac Patients on Coenzyme Q₁₀. Folkers K. & Yamamura Y. (eds.). pp. 281–290, Elsevier Science Publishers. Amsterdam.
- Petrović V. M., Saičić Z. S., Spasić M. B., Radojičić R. & Buzadžić B. (1991). Hormones and antioxidant defence, [In:] Anticarcinogenesis and Radiation Protection 2. Nygaard O. F. & Upton A. C. (eds.), pp. 405–416, Plenum Press, New York.
- Quiles J. L., Ramirez-Tortosa M. C., Huertas J. R., Ibanez S., Gomez J. A., Battino M. & Mataix J. (1999). Olive oil supplemented with vitamin E affects mitochondrial coenzyme Q levels in liver of rats after an oxidative stress induced by adriamycin. *BioFactors*, 9, 331–336.
- Saičić Z. S., Simović M., Korać B., Blagojević D., Buzadžić B., Spasić M. B. & Petrović V. M. (1993). Antioxidant defence system in erythrocytes and plasma of various strains of rats. Arch. Biol. Sci., 45, 89–92.
- Sarkar S., Yadav P. & Bhatnagar D. (1997). Cadmiuminduced lipid peroxidation and antioxidant system in rat erythrocytes: the role of antioxidants. J. Trace Elem. Med. Biol., 11, 8–13.
- Shukla G. S. & Chandra S. V. (1989). Cadmium toxicity and bioantioxidants: Status of vitamin E and ascorbic acid of selected organs in rat. J. Appl. Toxicol., 9, 119–122.
- Simpkins C., Lloyd T., Li S. & Balderman S. (1998).
 Metallothionein-induced increase in mitochondrial

- inner membrane permeability. J. Surg. Res., 75, 30 –34.
- Singhal R.K., Anderson M.E. & Meister A. (1987). Glutathione, a first line of defence against cadmium toxicity. FASEB J., 1, 220–223.
- Stadtman T.C. (1980). Selenium-dependent enzymes. Annu. Rev. Biochem., 49, 93-110.
- Takahashi T., Sugimoto N., Takahata K., Okamoto T. & Kishi T. (1996). Cellular antioxidant defence by a ubiquinol-regenerating system coupled with cytosolic NADPH-dependent ubiquinone reductase. Protective role against carbon tetrachloride-induced hepatotoxicity in the rat. Biol. Pharm. Bull., 19, 1005–1012.
- Tsuchihashi M. (1923). Zür Kenntnis der Blutkatalase. Biochem. Z., 140, 65–74.

- Waalkes M. P. & Oberdorster G. (1990). Cadmium carcinogenesis, [In:] *Metal Carcinogenesis*. Foulkes E. C. (ed.), pp. 845–852, CRC Press, Boca Raton.
- Yiin S. J., Chern C. L., Sheu J. Y., Tseng W. C. & Lin T. H. (1999). Cadmium-induced renal lipid peroxidation in rats and protection by selenium. J. Toxicol. Environm. Health, 57, 403–413.
- Zierz S., Jahns G. & Jerusalem F. (1989). Coenzyme Q in serum and muscle of 5 patients with Kearns-Sayre syndrome and 12 patients with ophtalmoplegia plus. J. Neurol., 236, 97–101.
- Žikić R. V., Štajn A. Š., Ognjanović B. I., Saičić Z. S.. Kostić M. M., Pavlović Z. S. & Petrović V. M. (1998). The effect of cadmium and selenium on the antioxidant enzyme activities in rat heart. J. Environm. Pathol. Toxicol. Oncol., 17, 259-264.